Study on drug susceptibility and bovine Hb using ELISA on bovine mastitis pathogens

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ABSTRACT:- Mastitis can be caused by a large variety of pathogens, some are infectious at the host level, and others are infectious at both host and population level. The aim of the present study was to to evaluate the drug sensitivity pattern of the isolated culture and Bovine milk Hp concentrations determination using a commercial bovine Hp ELISA kit. Milk samples were obtained with consent from the cows of different small holder dairy farms around Coimbatore District. The entire sample was collected according to the Scandinavian recommendation on examination of quarter milk samples and National Mastitis Council. The samples were cultured for the growth and isolation of microbes. Then the bacterial isolates subjected to biochemical tests for confirmation. The isolates were tested invitro for their antimicrobial susceptibility by agar disk diffusion method. Minimum inhibitory concentration (MIC) was analyzed for specific treatment and the Bovine Hp antibody used to determine the concentration of cell count presence in the milk sample.

Keywords:- Bovine mastitis, Bovine Hb, antibiotic resistance, MIC

I. INTRODUCTION

Mastitis is classified as a multi-etiological disease among the dairy animals which fetch considerable economic losses to the dairy industry throughout the world. Around 50 % of the herd populations of 70% of all available losses were observed in overall milk products (Sumathi et al., 2008). The factors which predispose cows to mastitis are presence of bacteria in and around the udder, poor hygiene, accumulation of milk, improper milking machines, breeds and injuries in teat are the factors which predispose cows to mastitis (Amehet al., 1993 and Egwuet al., 1999). The udder infection can be contracted at different ages and also at different stages of the lactation cycle. The ability to overcome an infection after the established of the same also varies (Klastrupet al., 1987). A broad spectrum of pathogens are identified as causatives for mastitis, the infection level can be classified as in the host level, population level and in some microorganism it is at both the level.

II. MATERIALS AND METHODS

The selection of dairy herds and aseptic sampling procedures will be followed according to the method described by Barkemaet al., 1998.

2.1 Milk Sample Collection

Milk samples were obtained with consent from the cows of different small holder dairy farms around Coimbatore District (Table-2.1). The entire sample was collected according to the Scandinivian recommendation on examination of quarter milk samples and National Mastitis Council (Medison, Laboratory Handbook on Bovine Mastitis, NMC, 1999). The protocol was permitted by the owners of the dairy farms under investigation. All efforts were made to minimize animal suffering.

| Table 2.1. Sampling places from Combatore Dist. | | | | | | | | | |
|---|-----------------|---|--------------------|--|--|--|--|--|--|
| S.No | Sampling Place | Latitude and Longitude | No. of Sample (SD) | | | | | | |
| 1 | Kannampalayam | 11 [°] 00'30.00"N 77 [°] 06'08.00"E | 12±0.8165 | | | | | | |
| 2 | Vellalur | 10°58'02.06"N 77°01'40.00"E | 10±0.5773 | | | | | | |
| 3 | Kodangipalayam | 10 [°] 47'27.06"N 77 [°] 02'08.08"E | 10±0.7071 | | | | | | |
| 4 | Karadivavi | 10 [°] 57'57.44"N 77 [°] 11'51.65"E | 15±1.2247 | | | | | | |
| 5 | Pappampatti | 10 [°] 57'34.30"N 77 [°] 06'06.38"E | 13±1.2583 | | | | | | |
| 6 | Malumichampatti | 10°54'06.32"N 76°59'45.55"E | 10±0.5773 | | | | | | |
| 7 | Saravanampatti | 11°05'24.00"N 76°59'52.00"E | 15±0.5477 | | | | | | |
| 8 | Somanur | 11°05'13.43"N 77°11'09.24"E | 12±1.1401 | | | | | | |
| 9 | Kinthukadavu | 10 [°] 49'00.00"N 77 [°] 01'00.00"E | 15±1.2247 | | | | | | |
| 10 | Pollachi | 10°39'17.52"N 76°59'22.27"E | 15±1.4142 | | | | | | |
| 11 | Mettupalayam | 11°17'57.14"N 76°55'29.26"E | 15±0.7071 | | | | | | |
| 12 | Karamadai | 11°14'31.68"N 76°57'10.09"E | 13±0.8165 | | | | | | |
| | | Total No. of Samples | 155±2.1087 | | | | | | |

| Table 2.1: | Sampling | places from | Coimbatore Dist. |
|------------|----------|-------------|-------------------------|
|------------|----------|-------------|-------------------------|

2.2 Identification of Bovine Mastitis Pathogen

The samples were cultured on Nutrient agar, Blood agar and MacConkey agar plates, supporting the growth of various types of bacteria for this study and isolation. The isolated bacteria were identified on the basis of their cultural and morphological characteristics. The pure cultures of bacterial isolates were obtained by sub culturing on differential and selective media. The bacterial isolates further subjected to biochemical tests for confirmation.

2.3 Antibiotics Sensitive Test

The isolates were tested invitro for their antimicrobial susceptibility by agar disk diffusion method in accordance with the standard in National Mastitis Council guidelines. Minimum inhibitory concentration (MIC) of 11 different antibiotics, ampicillin (10 μ g), Amoxicillin (10 μ g) Chloramphenicol (10 μ g), Gentamycin (10 μ g), Vancomycin (10 μ g), Tetracyclin (10 μ g), Ciprofloxacin (10 μ g), Ceftazidine (10 μ g), Cephalothin (10 μ g), Kanamycin (10 μ g), and streptomycin (10 μ g) was analyzed to suggest specific treatment.

2.4 Determination of milk Hp levels using ELISA

Freshly raw milk obtained from sampling places and immediately centrifuged. The top layer in the supernatant was carefully removed and remaining fractions (whey) used for Hp level analysis. Bovine milk Hp concentrations were determined using a commercial bovine Hp ELISA kit using ELISA, Thermo Fisher Scientific India Pvt. Ltd, India.(Chen et al.2006).

III. RESULTS AND DISSCUSSION

A total of 155 milk samples were collected for clinical and subclinical cases of mastitis from twelve different places in and around Coimbatore district. From each place, minimum of ten and maximum of 15 milk samples were collected. In all the milk samples, we have identified the presence of different pathogenic bacterias such as 240 number of E.coli sp., 431 of Staphylococcus aureus, 157 of Streptococcus, 321 of Bacillus sp., and40 of Y. enterocolitica sp. (Table-3.2). Major bacterial isolates were Staphylococcus aureus (36.2%), Bacillus sp. (27.0%), E.coli (20.2%), Streptococcus sp. (13.2%), and Yerseniaenterocolitica (3.4%) (Table-3.3). The maximum bacterial load identified from Vellalur, Karadivavi, Saravanampatti and Karamadai area samples. The Haptoglobin (Hp) is an acute phase protein responsive to inflammation and infection. One of the major functions of Hp is to capture released hemoglobin during excessive hemolysis. Even though Bovine Hp is not abundantly expressed in normal plasma but it is considered to be one of the sensitive acute phase proteins during bacterial infection. The present study was confirming the Bovine Mastitis microbial load in quantitative and qualitative manner. The Bovine Hp antibody used to determine the concentration of cell count presence in the milk sample by using ELISA. The high level of cells count presence in Karamadai 2.568±0.275µg/ml, Saravanampatti 2.506±0.375µg/ml, Karadivavi 2.284±0.290µg/ml and Vellalur 2.106±0.313µg/ml.(Table-3.4) We find an antibiotic resistant and sensitive strains from the entire microbial load. The antibiotic activity of gentamycin shown sensitivity of 96.2% and streptomycin was 94.5%. The mastitis pathogenic organism is highly sensitive to Gentamycin and Streptomycin where as other antibiotics are more resistant to E.coli and S.aureus (Table-3.5).

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| Milk sample- Coimbatore Dist. | | | | | | | | | | | |
|-------------------------------|----------------------|-----------------------|---------------|------------------|---------------|----------------------|--|--|--|--|--|
| | | | Staphylococcu | | | Y.enterocolitic a | | | | | |
| S.No | Sampling place | E.coli | S | Streptococcus | Bacillus sp. | | | | | | |
| | (No.of Sample) | (No.of Sample) (SD) a | | sp. (SD) | (SD) | (SD) | | | | | |
| 1 | Kannampalayam (12) | 12±0.127 | 23±0.735 | 17±0.628 | 19±1.783 | 04±0.100 | | | | | |
| 2 | Vellalur (10) | 18±0.352 | 48±1.008 | 14±0.332 | 38±1.654 | 07±0.121 | | | | | |
| 3 | Kodangipalayam (10) | 19±0.120 | 33±1.112 | 11±0.197 | 27±1.931 | Nil | | | | | |
| 4 | Karadivavi (15) | 25±0.187 | 52±0.937 | 14±0.188 | 40±2.101 | 09±0.137 | | | | | |
| 5 | Pappampatti (13) | 22±0.410 | 25±1.203 | 10±0.273 | 25±1.772 | 03±0.100 | | | | | |
| 6 | Malumichampatti (10) | 20±0.732 | 18±1.325 | 08±0.110 | 19±1.529 | Nil | | | | | |
| 7 | Saravanampatti (15) | 23±0.198 | 65±1.098 | 16±0.784 | 34±1.782 | 06±0.112 | | | | | |
| 8 | Somanur (12) | 20±0.110 | 27±0.999 | 19±0.931 | 27±1.900 | 04±0.120 | | | | | |
| 9 | Kinathukadavu (15) | 17±0.387 | 21±0.774 | 12±0.320 | 18±1.287 | 02±0.100 | | | | | |
| 10 | Pollachi (15) | 23±0.192 | 25±0.536 | 10±0.298 | 25±1.672 | Nil | | | | | |
| 11 | Mettupalayam (15) | 20±0.112 | 31±0.839 | 14±0.349 | 22±1.375 | Nil | | | | | |
| 12 | Karamadai (13) | 21±0.287 | 63±0.927 | 12±0.278 | 27±1.830 | 05±0.163 | | | | | |
| | Total | 240 | 431 | 157 | 321 | 40 | | | | | |

Table 3.2: Number of Isolation of Pathogenic bacterial species from Mastitis Milk sample- Coimbatore Dist.

Table 3.3: Total Number and Percentage of Bacterial isolates.

| S.No | Bacterial Species | No. of Isolates | % of isolates |
|------|------------------------|-----------------|---------------|
| 1. | E.coli | 240±3.384 | 20.2 |
| 2 | Staphylococcus aureus | 431±16.637 | 36.2 |
| 3 | Streptococcus sp | 157±3.203 | 13.2 |
| 4 | Bacillus sp | 321±7.262 | 27.0 |
| 5 | Yerseniaenterocolitica | 40±3.055 | 3.4 |
| | Total | 1189 | 100 |

Table-3.4: Haptoglobin (Hp) concentration of isolated Bovine milk samples.

| S.No | Sampling Places | No. of Sample | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | Total (S.D) |
|------|---------------------|------------------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|---------------|-----------|-----------|-----------------|
| 1 | Kannampala yam | 12 | 0.8 31 | 0.6 38 | 0.7 42 | 0.83 7 | 0.8 01 | 0.4 71 | 0.5 24 | 0.2 36 | 0.81 1 | 0.7 28 | 0.6 71 | 0.6 49 | eπe | Ţ | Ţ | 0.662 ±0.178 |
| 2 | Vellalur | 10 | 1.7 19 | 1.6 93 | 2.7 14 | 2.32 5 | 1.9 98 | 2.1 24 | 1.8 37 | 2.1 12 | 2.31 | 2.2 25 | 17 | - | 8.0 | 7 | Ξ | 2.106±0.3 13 |
| 3 | Kodangipala yam | 10 | 0.2 37 | 0.3 15 | 0.2 16 | 0.29 9 | 0.3 12 | 0.3 27 | 0.2 86 | 0.2 74 | 0.30 1 | 0.2 78 | | <u> 1</u> | anne. | an. | | 0.285±0.0 35 |
| 4 | Karadivavi | 15 | 1.8 97 | 2.0 13 | 2.5 64 | 2.32 7 | 2.7 84 | 2.3 27 | 2.4 28 | 2.3 33 | 2.56 9 | 1.8 72 | 1.9 77 | 2.3 14 | 2.71 | 2.6 25 | 2.5 42 | 2.284±0.2 90 |
| 5 | Pappampatti | 13 | 0.5 42 | 0.6 16 | 0.4 71 | 0.52 9 | 0.5 87 | 0.6 11 | 0.6 27 | 0.5 88 | 0.42 1 | 0.4 39 | 0.4 98 | 0.5 2 | 0.39 9 | - | - | 0.537±0.0 70 |
| 6 | Malumicham patti | 10 | 0.3 | 0.2 99 | 0.2 71 | 0.28 9 | 0.3 11 | 0.3 02 | 0.2 77 | 0.2 78 | 0.31 4 | 0.2 89 | = | - | (<u>4</u> 4) | 4 | = | 0.295±0.0 17 |
| 7 | Saravanamp atti | 15 | 1.9 24 | 1.7 35 | 2.9 01 | 2.87 3 | 2.8 01 | 2.5 42 | 2.4 21 | 2.3 38 | 2.36 | 2.5 93 | 2.7 09 | 2.8 72 | 2.63 8 | 2.4 79 | 2.1 18 | 2.506±0.3 75 |
| 8 | Somanur | 12 | 0.3 | 0.3 64 | 0.3 | 0.32 | 0.3 16 | 0.2 97 | 0.2 67 | 0.3 11 | 0.30 | 0.3 | 0.3 28 | 0.2 89 | <u> </u> | 7 | Ξ | 0.320±0.0 33 |
| 9 | Kinathukada vu | 15 | 0.2 25 | 0.2 29 | 0.2 83 | 0.25 5 | 0.2 67 | 0.2 91 | 0.2 83 | 0.3 11 | 0.22 1 | 0.1 97 | 0.2 73 | 0.2 99 | 0.27 5 | 0.3 01 | 0.2 69 | 0.261±0.0 36 |
| 10 | Pollachi | 15 | 0.5 44 | 0.5 21 | 0.6 32 | 0.59 8 | 0.5 62 | 0.6 11 | 0.6 18 | 0.6 15 | 0.47 1 | 0.4 79 | 0.3 28 | 0.3 79 | 0.42 8 | 0.4 73 | 0.4 59 | 0.530±0.0 99 |
| 11 | Mettupalaya m | 15 | 0.2 25 | 0.2 74 | 0.2 93 | 0.42 8 | 0.3 99 | 0.4 21 | 0.4 01 | 0.2 97 | 0.28 3 | 0.2 94 | 0.4 22 | 0.3 85 | 0.36 | 0.3 99 | 0.3 43 | 0.344±0.0 72 |
| 12 | Karamadai | 13 | 1.9 87 | 2.3 42 | 2.9 34 | 2.99 1 | 2.7 32 | 2.5 43 | 2.4 63 | 2.7 25 | 2.57 7 | 2.6 32 | 2.5 78 | 2.3 14 | 2.71 4 | 2.6 34 | 2.5 81 | 2.568±0.2 75 |

REFERENCES

- [1] Ameh, J.A., Addo, P.B., Adekeye, J.O., and Gyang, E.O., Prevalence of clinical mastitis and of intermammary infection in Nigeria goats, Preventive Veterinary Medicine 17 (1-2), 1993, 41-46.
- [2] Barkema HW, Westrik JD, van Keulen KAS, Schukken YH, Brand A, The effects of lameness on reproductive performance, milk production and culling in Dutch dairy herds. Preventive Veterinary Medicine 20, 1994, 249-259.
- [3] Chen, B.S. and Wang, Y.C, On the attenuation and amplification of molecular noise in genetic regulatory networks, BMC bioinformatics, 2006, 7:52.
- [4] Egwu, G. O., J. A. Ameh, M. M. Aliyu, F. D. Mohammed, Caprinemycoplasmal mastitis in Nigeria, Vet. arhiv 69, 1999, 241-250.
- [5] Klastrup O., Bakken G., Bramley J., Bushnel R, Environmental influences on bovine mastitis, Bulletin of the International Dairy Federation, 217, 1987, 37.
- [6] Laboratory Handbookon Bovine MastitisPublished by theNational Mastitis Council
- [7] Sumathi, B.R., Veeregowda, B.M., and Amitha, R. Gomes, Prevalence and antibiogram profile of bacterial isolates from clinical bovine mastitis, Veterinary World,2008, 1, 8.